

# High Expression of Heat Shock Protein Family D Member 1 Predicts Poor Prognosis of Esophageal Cancer

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## Abstract

**Background:** Heat shock protein family D (Hsp60) member 1 (HSPD1) has been reported as a potential survival-related biomarker in some cancers. However, the correlation between HSPD1 expression with prognosis and clinical features of esophageal cancer (EC) is poorly understood. Our research aimed to explore the clinical and prognostic significance of HSPD1 expression in EC patients.

**Methods:** In our study, HSPD1 expression was detected by immunohistochemistry in 87 EC tissue specimens and 20 normal cancerous peripheral tissue specimens. Meanwhile, we also analyzed the expression of HSPD1 in EC by The Cancer Genome Atlas (TCGA) database. Then Chi-squared and Fisher's exact tests and Wilcoxon signed-rank test and logistic regression models were separately used to test the correlation between clinical characteristics and HSPD1 expression in our and TCGA cohort. Moreover, we evaluated the value of HSPD1 in prognosis by Kaplan-Meier curves and Cox analysis. Finally, gene set enrichment analysis (GSEA) was performed using the data accessed from TCGA.

**Results:** The results showed that HSPD1 was overexpressed in EC, and the expression was related to histological type, histological grade, N classification, and clinical stage. Moreover, Kaplan-Meier curves and Cox analysis indicated that high expression of HSPD1 correlated with poor prognosis, and HSPD1 was an independent risk factor for EC. GSEA identified pathways involved in cysteine and methionine metabolism, spliceosome, selenoamino acid metabolism, mismatch repair, RNA degradation, DNA replication, and cell cycle as differentially enriched in ECs with high HSPD1 expression.

**Conclusions:** Our results suggest that HSPD1 is expressed at high levels in EC, and has potential to be used as a novel biomarker for the prognosis of patients with EC.

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**Keywords:** HSPD1; Esophageal cancer; Prognosis

## Introduction

Esophageal cancer (EC) is the ninth most common cancer and sixth most common cause of cancer-related death globally [1]. Every year more than 400,000 people suffer from EC and the incidence rates are increasing rapidly [2], with about 40% of that in China alone [3]. Although the prognosis and survival have improved, the 5-year survival rate remains low [4]. Therefore, it is essential to find more novel potential prognostic biomarkers for improving the prognoses of EC patients.

Heat shock proteins (HSPs) are groups of genetically highly conserved proteins involved in maintaining cell homeostasis during normal physiology [5]. According to the different molecular weights, these proteins have been classified, including HSPB1 (HSP27), DNAJB1 (HSP40), HSPD1 (HSP60), HSPA4 (HSP70), HSP90AA1 (HSP90) and HSPH (HSP110) [6]. Besides their cytoprotective effects, previous studies have demonstrated HSPs involved in the development, progression, metastasis and drug resistance of cancers [7]. Potential clinical roles of some HSPs in ECs have been reported. For example, Hsp27 activation increased ALDH activity, chemoresistance and tumor initiation in esophageal squamous cell carcinoma (ESCC) cell lines, and is considered as a prognostic indicator in ESCC [8, 9]. High expression of HSP47 is associated with poor prognosis in patients with ESCC [10]. HSP90a has potential clinical application as a predictor of response to chemoradiotherapy, and might be an independent prognostic factor for ESCC [11, 12]. However, the clinical significance of HSPD1 in EC is still not clear. Therefore, our research aimed to explore the clinicopathological significance of HSPD1 expression in EC patients by a combined method of immunohistochemistry and bioinformatics.

## Materials and Methods

### Tumor specimens and clinical data collection

We retrospectively searched our institutional database for EC patients between January 2013 and December 2017. Patients who underwent chemotherapy and/or radiotherapy prior to

surgery or biopsy were excluded from the study. Then, a total of 87 paraffin-embedded EC tissue specimens and 20 adjacent non-neoplastic tissue specimens were collected. The pathological diagnosis of each tissue specimen was confirmed by at least two pathologists. Medical records of each patient were used to extract data including age, gender, histological type, histological grade, stage classification, T classification, N classification and M classification. The date of diagnosis was set as the starting point and the date of death or last date of follow-up as the end point. The present study was approved by the Ethics Committee of People's Hospital of Deyang City (Deyang, China) and all human tissue samples were obtained following written informed consent.

### Immunohistochemistry

Immunohistochemical staining with HSPD1 (1:100 dilution; Abcam) was accomplished using Dako Link 48 autostainer (DAKO) following the manufacturer's instructions. Immunostaining was evaluated independently by two senior pathologists who blinded to the clinical data, on at least 10 random fields at 400 magnification and cytoplasmic stained cells were counted. The immunohistochemical expression was scored as follows: high expression, > 50% and low expression, < 50% of the cancer cells positive staining.

### Data collection from The Cancer Genome Atlas (TCGA) database

The data of EC patients and mRNA expression profiles (169 cases, including 10 normal samples) was downloaded from the TCGA database (<https://cancergenome.nih.gov/>). The expression difference of HSPD1 was shown as a box plot. P values below 0.05 were considered statistically significant. The associations between clinical features and HSPD1 expression were assessed by the Wilcoxon signed-rank test and logistic regression.

### Gene set enrichment analysis (GSEA)

In this study, GSEA was performed to find the significant pathways between low expression and high expression group of HSPD1 by using the GSEA software (4.1.0). The high and low expression groups were defined as the median value of expression level of HSPD1. The normalized enrichment score (NES) was acquired by investigating permutations for 1,000 times. A gene set is considered to signify the statistical significance when a normal P-value is < 0.05 as well as false discovery rate (FDR) is < 0.05. The graphical results are shown in one diagram by using R software (V.4.0.2).

### Statistical analysis

The relationships between HSPD1 expression and clinico-

pathological features were analyzed using the Chi-square test or Fisher's exact test in our study. Kaplan-Meier curves were performed, and the differences in the overall survival were compared by log-rank test. Univariate Cox analysis was conducted to select the significant related parameters. Then, the multivariate Cox analysis was applied to assess the independent prognostic factor for overall survival of EC patients. The result of multivariate Cox analysis was shown as a forest plot by using the *survminer* R package. All statistical analyses were performed using R software (V.4.0.2).

## Results

### High HSPD1 expression in EC

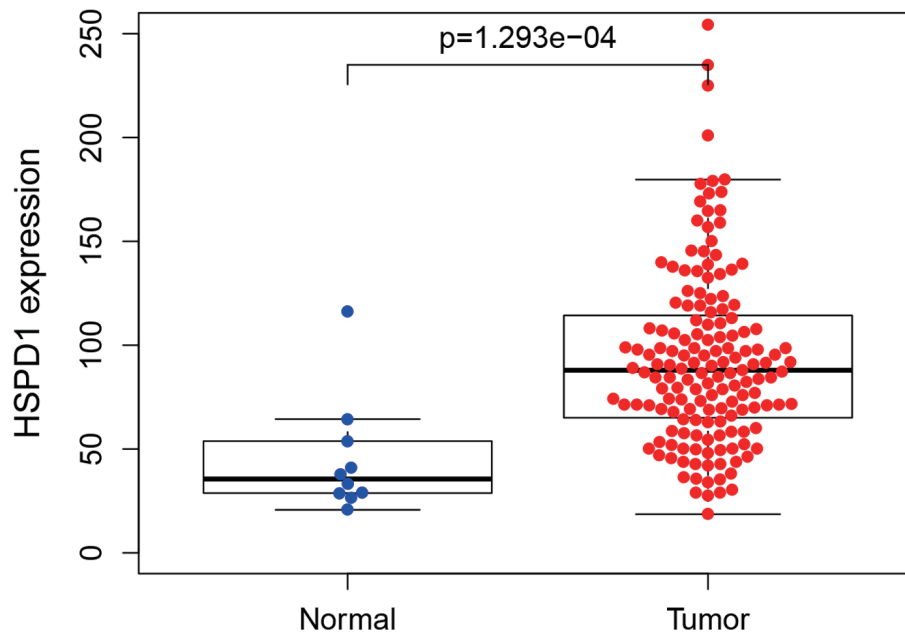
In total, 169 EC and adjacent non-neoplastic tissue samples from the TCGA database were included in the current comparison. As shown in Figure 1, the HSPD1 expression was elevated in EC tissues compared with adjacent non-neoplastic tissues ( $P = 0.0237$ ). In our study, we performed immunohistochemical analysis to assess HSPD1 expression in EC tissues and normal adjacent non-neoplastic tissues (Fig. 2). HSPD1 was mainly expressed in the cytoplasm. In the EC tissues, high expression of HSPD1 was observed in 54% (47/87) tumor samples. While in the adjacent non-neoplastic tissues, 10% (2/20) of samples exhibited high HSPD1 expression. The statistical result suggested that the expression of HSPD1 in EC tissues was higher ( $P < 0.001$ , Table 1), which was consistent with the results of TCGA database.

### Associations between HSPD1 expression and clinical features in EC patients

In TCGA cohort, the connections between the clinicopathological characteristics and the expression of HSPD1 were analyzed and summarized in Figure 3 and Table 2. HSPD1 expression was notably related with histological type ( $P = 0.001$ ), clinical stage ( $P = 0.024$ ), T classification ( $P = 0.003$ ), N classification ( $P = 0.007$ ) and M classification ( $P = 0.008$ ) (Fig. 3). Univariate analysis using logistic regression showed that high expression of HSPD1 in EC was significantly associated with high N classification (odds ratio (OR) = 2.109 for N0 vs. N1-3), adenocarcinoma pathological type (OR = 0.488 for adenocarcinoma vs. squamous carcinoma). In our study, the expression of HSPD1 was highly associated with the clinical stage (I-II vs. III-IV,  $P = 0.001$ , Table 3), N classification (N0 vs. N1-3,  $P = 0.014$ , Table 3) and histological grade (well vs. moderately/poorly,  $P < 0.001$ , Table 3).

### Expression of HSPD1 is associated with the overall survival

Firstly, Kaplan-Meier survival analysis along with the log-rank test was performed based on expression level of HSPD1 in EC cohort from TCGA database, and we found that EC patients with high HSPD1 expression were closely associated with



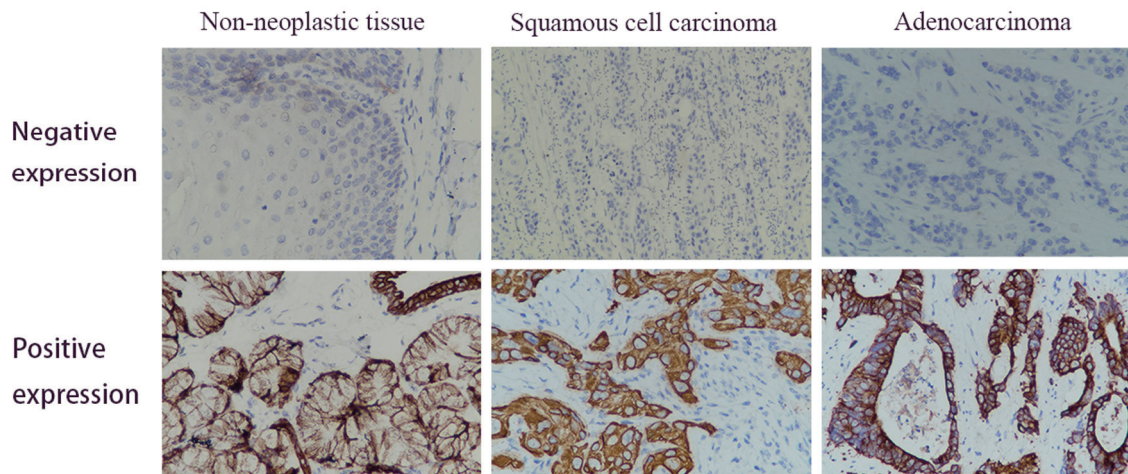
**Figure 1.** Expression of HSPD1 in esophagus cancer patients based on TCGA data. The expression level of HSPD1 in esophagus cancer tissues was significantly higher than that in adjacent non-neoplastic tissues ( $P = 0.0237$ ). HSPD1: heat shock protein family D (Hsp60) member 1; TCGA: The Cancer Genome Atlas.

poor overall survival than those with a low HSPD1 expression ( $P = 0.012$ ; Fig. 4a). This relationship was validated in the EC cohort from our study ( $P = 0.008$ , Fig. 4b). Secondly, high HSPD1 expression, clinical stage, N classification and M classification were selected by univariate Cox proportional hazards regression analysis. Furthermore, the result of multivariate Cox proportional hazards regression analysis suggested that high HSPD1 expression (hazard ratio (HR) = 1.572) and advanced clinical stage (HR = 2.281) were independent prognostic factors for EC patients in TCGA cohort (Fig. 5a). These findings were consistent with the results based on our EC co-

hort (HR = 1.595 for high HSPD1 expression and HR = 1.679 for advanced clinical stage, Fig. 5b).

#### GSEA identifies HSPD1-associated signaling pathways

In order to explore potential biological pathways that were activated in different groups, GSEA was performed. GSEA revealed significant differences in the enrichment of the MSigDB collection (c2\_cp.kegg.v7.1.symbols.gmt). As shown in Figure 6, gene sets related to cysteine and methionine metabolism,

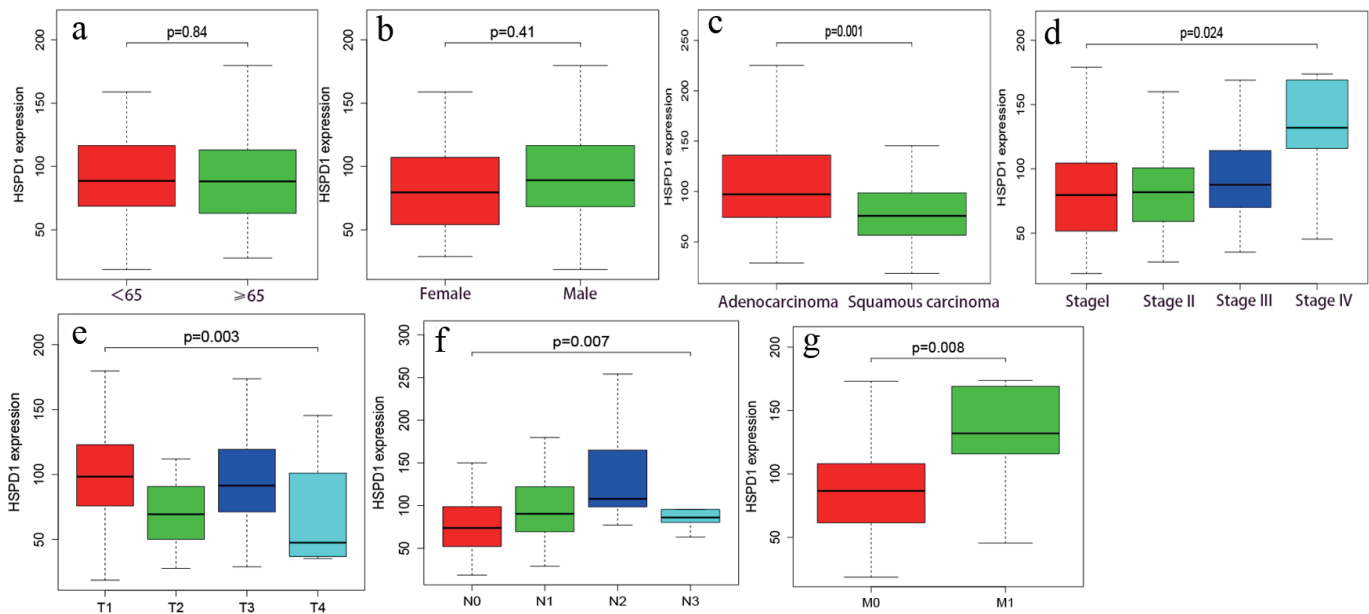


**Figure 2.** Immunohistochemical analysis of the HSPD1 expression in esophagus cancer and adjacent non-neoplastic tissues. Positive expression of HSPD1 was mainly found in the cytoplasm (magnification,  $\times 400$ ). HSPD1: heat shock protein family D (Hsp60) member 1.

**Table 1.** HSPD1 Expression in Esophagus Cancer Tissues and Adjacent Non-Neoplastic Tissues

Group	n	HSPD1 expression		P value
		Low (%)	High (%)	
Normal	20	18 (90%)	2 (10%)	< 0.001*
Tumor	87	40 (46%)	47 (54%)	

\*P < 0.05. HSPD1: heat shock protein family D (Hsp60) member 1.



**Figure 3.** Association of HSPD1 expression with clinical variables based on TCGA data. (a) Age. (b) Gender. (c) Histological type. (d) Clinical stage. (e) T classification. (f) N classification. (g) M classification. HSPD1: heat shock protein family D (Hsp60) member 1; TCGA: The Cancer Genome Atlas.

**Table 2.** Logistic Regression of HSPD1 Expression and Clinicopathological Characteristics in TCGA Database

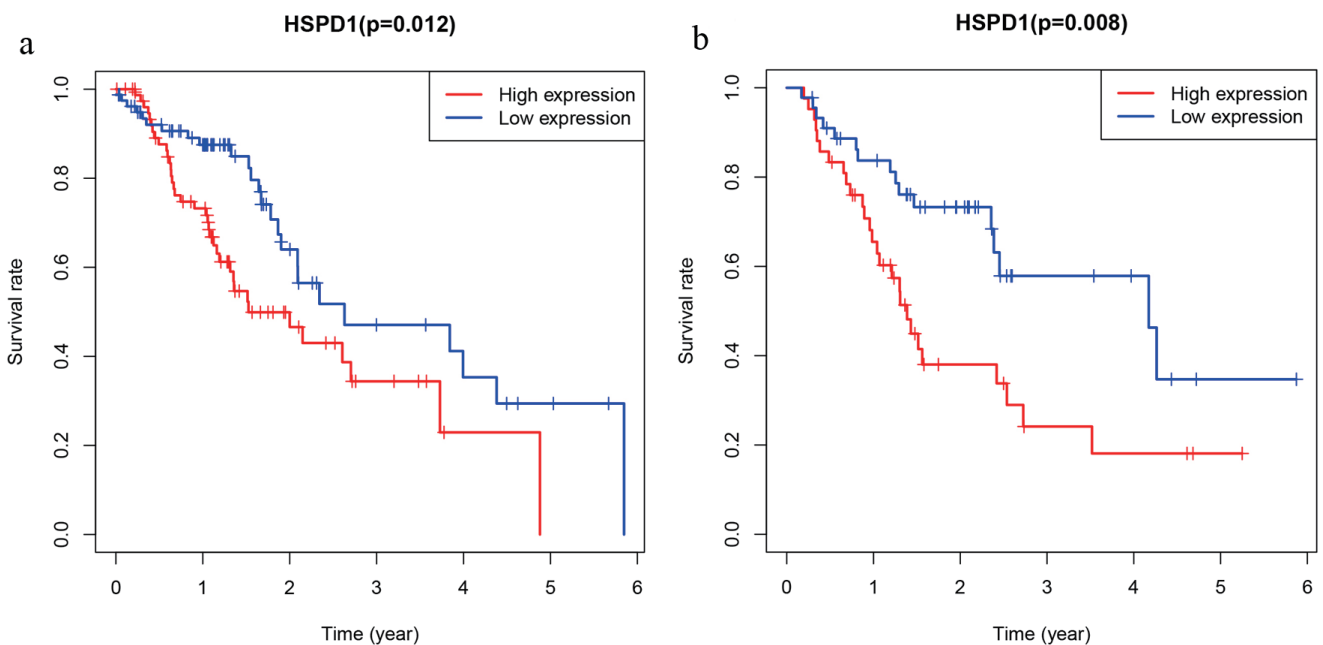
Clinical characteristics	n	Odds ratio in expression	P value
Age			
< 65 vs. ≥ 65	159	0.995 (0.520 - 1.92)	0.893
Gender			
Female vs. male	159	1.359 (0.56 - 3.39)	0.45
Pathology type			
Adenocarcinoma vs. squamous carcinoma	158	0.488 (0.257 - 0.916)	0.027*
Stage			
I-II vs. III-IV	140	1.823 (0.923 - 3.646)	0.086
T			
T0-2 vs. T3-4	144	1.863 (0.962 - 3.648)	0.067
N			
N0 vs. N1-3	142	2.109 (1.083 - 4.167)	0.029*
M			
M0 vs. M1	128	7.875 (1.343 - 149.581)	0.057

\*P < 0.05. HSPD1: heat shock protein family D (Hsp60) member 1; TCGA: The Cancer Genome Atlas.

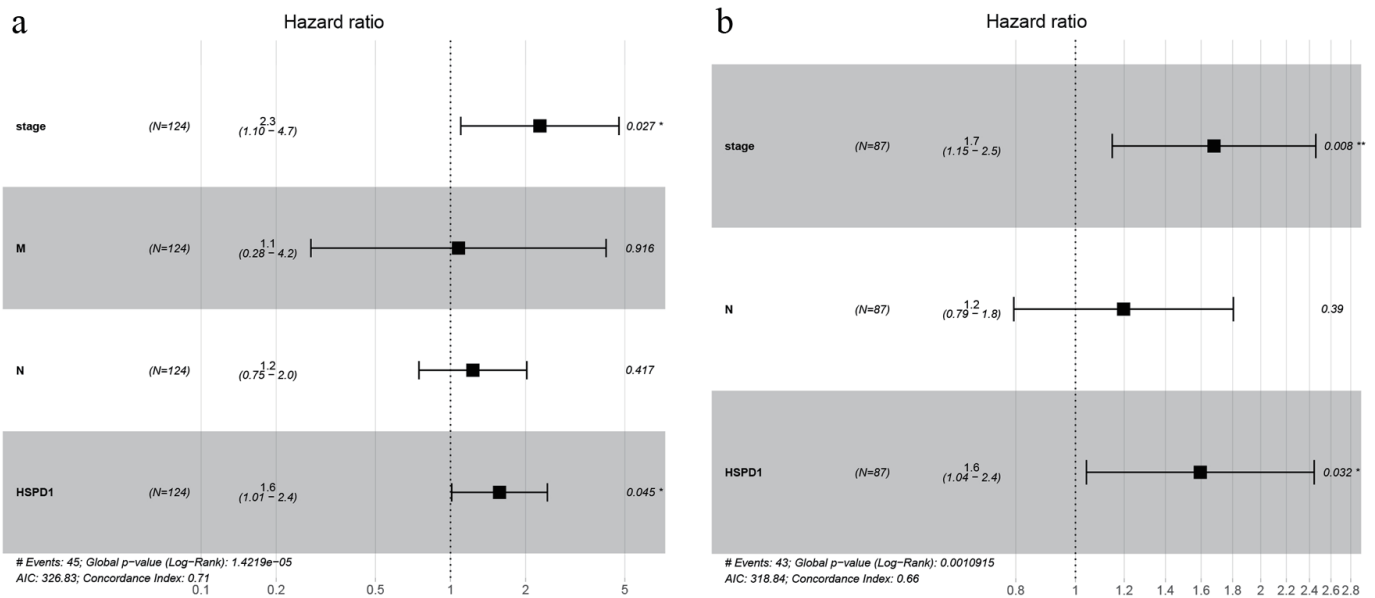
**Table 3.** Associations Between HSPD1 Expression and Clinicopathological Characteristics in Our Esophagus Cancer Patients

Characteristics	n	HSPD1 protein expression		P value
		Low (%)	High (%)	
Age				0.996
≥ 65	50	23 (46.0)	27 (54.0)	
< 65	37	17 (45.9)	20 (54.1)	
Stage				0.001*
I-II	42	27 (64.3)	15 (35.7)	
III-IV	45	13 (28.9)	32 (71.1)	
T				0.381
T0-T2	50	25 (50.0)	25 (50.0)	
T3-T4	37	15 (40.5)	22 (59.5)	
N				0.014*
N0	42	25 (59.5)	17 (40.5)	
N1-N3	45	15 (33.3)	30 (66.7)	
M				0.335
M0	80	38 (47.5)	42 (52.5)	
M1	7	2 (28.6)	5 (71.4)	
Histological type				0.116
Squamous carcinoma	75	37 (49.3)	38 (50.7)	
Adenocarcinoma	12	3 (25.0)	9 (75.0)	
Histological grade				< 0.001*
Well	40	27 (67.5)	13 (32.5)	
Moderately/poorly	47	13 (27.7)	34 (72.3)	

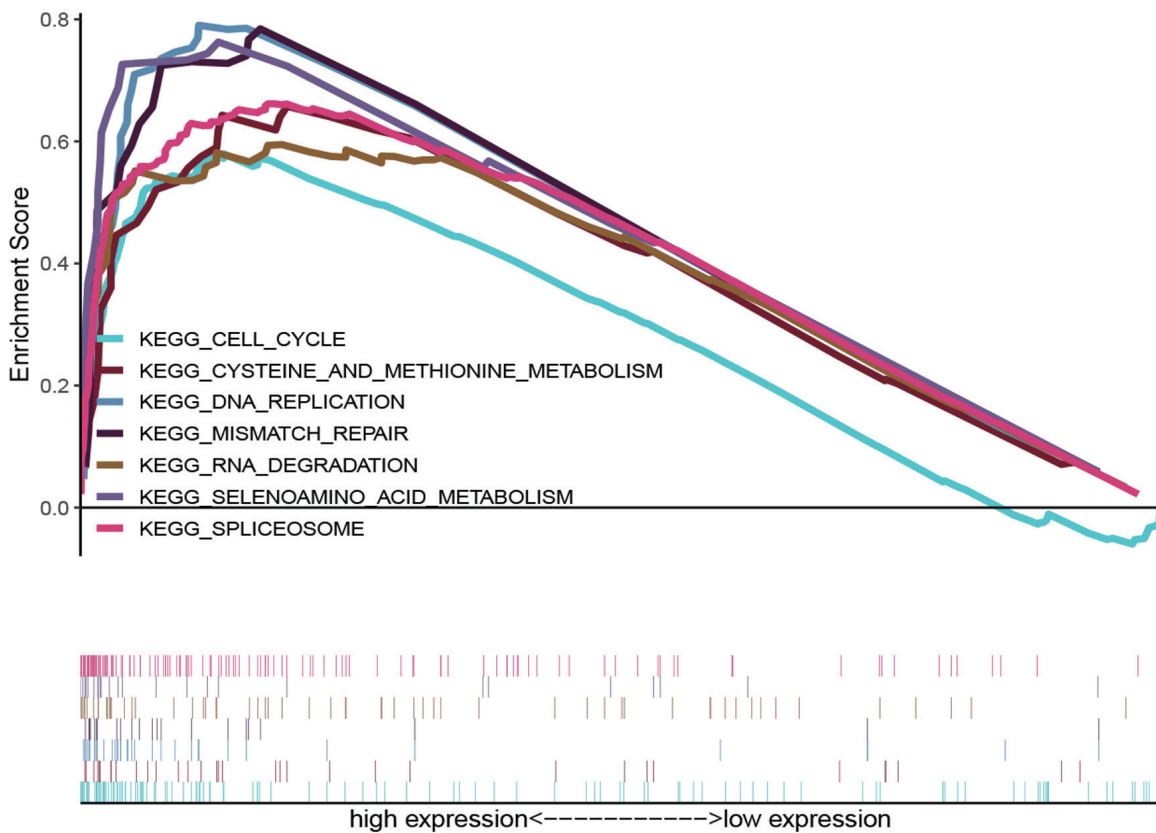
\*P < 0.05. HSPD1: heat shock protein family D (Hsp60) member 1.



**Figure 4.** The prognostic significance of HSPD1 in esophagus cancer. Kaplan-Meier method and log-rank test were performed based on expression level of HSPD1 in esophagus cancer cohort from TCGA database (a) and our study (b). HSPD1: heat shock protein family D (Hsp60) member 1; TCGA: The Cancer Genome Atlas.



**Figure 5.** Forest plot of multivariate Cox regression analyses of overall survival in esophagus cancer cohort from TCGA database (a) and our study (b). TCGA: The Cancer Genome Atlas.



**Figure 6.** KEGG enrichment plots from GSEA. The GSEA results revealed that genes involved in cysteine and methionine metabolism, spliceosome, selenoamino acid metabolism, mismatch repair, RNA degradation, DNA replication and cell cycle were differentially enriched in HSPD1-associated esophagus cancer. HSPD1: heat shock protein family D (Hsp60) member 1; GSEA: gene set enrichment analysis.

spliceosome, selenoamino acid metabolism, RNA degradation, cell cycle, mismatch repair and DNA replication were associated with the HSPD1 high expression phenotype.

## Discussion

HSPD1 is a nuclear-encoded mitochondrial protein, primarily but not exclusively localized in the mitochondrial matrix [13]. Together with co-chaperonin HSP10, it facilitates correct folding and assembly of imported proteins in the mitochondria, and as a signaling molecule that activates a class of immunoreaction [14]. Besides, it has pro-inflammatory functions, and plays a role of dual pro-survival [15, 16] and pro-apoptosis [17, 18] functions. Recently, HSPD1 has been considered as a prognostic biomarker for poor overall survival involved in several types of cancers [19-21]. However, in EC, the relationship between HSPD1 expression and tumor histopathology or clinical prognosis is controversial. In an early study, Faried et al [22] reported that positive HSPD1 expression in ESCC contributed to the induction of apoptosis and correlated with favorable prognosis. In contrast, in a study from China, expression of HSPD1 was significantly increased in ESCC, and associated with poor disease survival [23]. Therefore, the correlation between HSPD1 and EC remains to be further explored.

HSPD1 was expressed at markedly higher levels in several kinds of cancers, such as oral [20], prostate [24], gastric [19], colorectal [25] or cervical [26] cancer. However, low HSPD1 expression was observed in ovarian cancer [27]. In the present study, we found that EC tissues exhibited high HSPD1 expression in comparison with adjacent non-neoplastic tissue, which was consistent with the expression status of HSPD1 in TCGA database. Together with our study in EC, these results suggested that HSPD1 expression levels may be associated with tumorigenesis in most types of human cancer. Further, we found that HSPD1 expression was markedly associated with advanced clinical stage, more lymph node metastasis and worse histological grade. The result of univariate analysis using logistic regression models suggested high expression of HSPD1 was significantly associated with high N classification and adenocarcinoma pathology type in TCGA cohort, which was partly consistent with our findings. We further evaluated the association between HSPD1 expression and overall survival of EC patients in TCGA database, and found HSPD1 expression was negatively correlated with overall survival time in EC patients. Furthermore, similar result has shown in the overall survival curve of our cohort. Univariate and multivariate Cox proportional hazards regression analyses of both TCGA database and our cohort indicated HSPD1 expression was an independent factor for poor prognosis in EC patients. Generally, high HSPD1 expression is a credible biomarker for predicting poor prognosis in EC patients.

Recently, HSPD1 has been found in many extramitochondrial sites, including the extracellular surface, cell surface, intracellular vesicles, nucleus, extracellular fluid, and even the cytoplasm [28]. It has been reported that HSPD1, especially which consists in cytosolic, is involved in an increased ability of metastasis and cell survival of various cancers [19, 21,

29]. Tsai et al [30] reported that cytosolic HSPD1 interacts with  $\beta$ -catenin to promote metastasis through enhancing transcriptional activity and increasing protein levels of  $\beta$ -catenin in head and neck cancer. Moreover, it has been reported that HSPD1 represses E-cadherin expression at transcriptional and translational levels through RelA activation and may contributes to metastasis in buccal mucosa squamous cell carcinoma (BMSCC) cells [20]. Downregulation of HSPD1 could induce the apoptosis of gastric cancer cells and was negatively correlated with the MEK/ERK signaling *in vitro* [31]. Also, inhibition of HSPD1 could suppress the proliferation of glioblastoma cells through the ROS/AMPK/mTOR pathway [32]. Although many potential functions have been presented for HSPD1 in various types of cancer, no report relates HSPD1 to biological role in EC so far. To further evaluate the roles of HSPD1 in EC, we performed GSEA using TCGA data. GSEA showed that genes involved in cysteine and methionine metabolism, spliceosome, selenoamino acid metabolism, RNA degradation, cell cycle, mismatch repair and DNA replication were associated with the HSPD1 high expression phenotype. It has been reported that methionine levels might be vital for promoting proliferation and drug resistance of cancers [33, 34]. Moreover, methionine deprivation can stop tumors from growing in various cancers, and chemo-sensitize cancer cells [35-38]. A recent study showed that H<sub>2</sub>S-producing enzyme cystathionine  $\gamma$ -lyase (CTH) which was involved in the metabolism of cysteine and methionine could generate H<sub>2</sub>S to promote prostate cancer metastasis and progression by IL-1 $\beta$ /NF- $\kappa$ B signaling pathways [39]. To date, several studies have reported that alternative splicing provides the potential to generate diversity at RNA and protein levels from an apparently limited number of loci in the genome. Dysregulation of alternative splicing characterizes many cancers and is sufficient to drive disease initiation, progression, and therapeutic response [40-42]. Moreover, mismatch repair capacity, RNA degradation, DNA replication and cell cycle pathway are also the critical mechanism in cancer progression [43-46]. Although we have demonstrated that the value of HSPD1 expression is a potential reliable molecular marker for the prognosis in EC, some limitations of our study should be noted. First, the sample used in this research is limited. Second, further exploration needs to be carried out in the future to verify the detailed molecular mechanisms of HSPD1 expression in EC.

## Conclusions

HSPD1 expression is up-regulated in EC tissues, and positively associated with clinical progression in EC patients. In patients with EC, the expression levels of HSPD1 and clinical stage were independent prognostic factors. Further studies should be performed to evaluate the diagnostic and prognostic role of HSPD1 and its potential molecular role as a therapeutic target.

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None to declare.

## Financial Disclosure

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## Conflict of Interest

The authors declare that they have no competing interests.

## Informed Consent

Not applicable.

## Author Contributions

PC, WX, XS, JY, and JL performed the experiments; WX performed the statistical analysis; and PC designed the study, analyzed the data and drafted the manuscript. All authors read and approved the final manuscript.

## Data Availability

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

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